

## บทบาทของ DNA microarray ในโรคติดเชื้อ

กนิษฐา ภัทรกุล\*

DNA microarrays or gene chips are developed based on the currently available information of complete sequences of human and several pathogen genomes and the technology that enables placing of nucleic acid probes on a single array at a microscopic and high-density scale.<sup>(1)</sup> The working principle behind DNA microarrays is parallel to traditional Southern hybridization, its "macro" array counterpart. The novel fabrication technologies of DNA microarray can immobilize tens of thousands genes (or oligonucleotides) or all genes of an entire genome on a solid substrate or platform in comparison to hundreds of genes on Southern blot.<sup>(2-4)</sup> Samples for microarrays can either be microbial DNA content or their gene-expression profiles. Hybridization of labeled targets derived from nucleic acids in the test sample to the probes on the array makes probing multiple gene targets possible in a single experiment. Thus, DNA microarray is a high-throughput technology for detection and analysis of genes and gene expression on a "global" or "genome-wide" scale.

Since its emerging in the early 1990s, the number of studies using DNA microarrays has risen markedly during the past decade.<sup>(3,5)</sup> Early microarray studies focused mainly on human cancers due to the readily available tumor specimens and the social

impact of the diseases.<sup>(6-8)</sup> DNA microarray technology has been gradually becoming a standard tool in regular biological laboratories. This tool is presently applied to other fields of medicine including the infectious diseases.<sup>(9-11)</sup> The potential of the DNA microarrays in investigation of both sides of the host-pathogen interaction offers a mean to study infectious diseases in a systemic and efficient way.<sup>(12-14)</sup>

A large number of DNA sequences including antigenic determinants, virulence factors, and multiple different sequence variants of each gene target immobilized on the microarrays allow the detection of a broad range of pathogens with high specificity and discriminatory ability.<sup>(15)</sup> Therefore, identification of pathogens via DNA microarrays needs not be restricted to a single strain, species, genus, or class of organisms. This advantage is beneficial for identification of causative agents of infectious diseases. Although detection of microbes using the currently available molecular techniques is faster than conventional methods, discrimination of very closely related species or different organisms without prior amplification of target DNA or pre-identification of the pathogen remains difficult. Several studies demonstrated the efficiency of DNA microarrays for rapid and simultaneous identification of closely related

pathogens and/or distinct etiologic organisms in a single assay.<sup>(16-27)</sup> In addition, depending on probes designed on the platform DNA microarrays have been previously shown to be a tool with broad identification capacity to identify virulence factors, detect polymorphisms or mutation in the pathogens, study complex microbial populations, determine taxonomic relationships between microbial strains at species to strain level resolution, determine molecular typing, identify multiple antibiotic resistance genes, survey the spread of strain or antibiotic resistance determinants in epidemiological studies, and discover unknown causative agents of infectious diseases.<sup>(24, 28-40)</sup>

The crucial advantage of microarray-based approach allows the entire biological pathways and coordinated interaction of multiple pathways or genes to be studied without prior bias to a particular gene or pathway. Thus, microarrays have been used for genome-wide analysis of host expression responses to diverse pathogenic stimuli and differential gene expression of microbial virulence factors during infection providing an insight of the host-pathogen interaction, the key to pathogenesis of infectious diseases.<sup>(14, 41, 42)</sup> To study host immune responses, global expression analyses of both the innate and adaptive immune cells at various stages of differentiation, maturation and activation during microbial invasion is an unbiased approach to comprehend the complex coordination of immune responses to infection.<sup>(9)</sup> Numerous studies have utilized microarray analysis to follow gene expression alterations of host cells, either target cells or immune cells, in response to interactions with infectious pathogens, and vice versa.<sup>(43-55)</sup> Distinctive host gene

expression patterns observed in patients infected with different etiologies might assist in the differential diagnosis of infectious diseases.<sup>(56)</sup> The relationship between expression profiles of host inflammatory cells and clinical status may be useful to predict clinical outcome.<sup>(57)</sup> In addition, susceptibility to infectious diseases and clinical response to antimicrobial drugs may be associated to certain host genetic background.<sup>(58, 59)</sup> Thus, global analysis of host and microbial interaction contributes to new insights on pathogenesis of infectious diseases leading to novel strategies for therapeutic and prophylactic interventions and prognostic markers of outcome.<sup>(11, 60-64)</sup>

Microarrays also have limitation or disadvantages. Although the DNA microarray technology is a genome-wide approach, the detection is restricted to DNA immobilized on the platform.<sup>(9)</sup> As a result, this technique cannot detect sequences or genes that are absent on the array. The genes or oligonucleotides on the array need to be updated according to currently available information. Like other molecular methods, standardization of the system is required before the microarray technique is implemented for clinical practice in infectious diseases.<sup>(65)</sup> Due to its early application, protocols are not well-defined. In addition, the cost of equipments and software is still high. Personnel require special training and complementary knowledge. Mathematical and statistical knowledge and/or experts are necessary for study design and data analysis due to substantial amount of data derived from this technique. Therefore, most microarray techniques are presently used in research.

In author's opinion, the main advantage of DNA microarray technique is its rapid, sensitive and high-throughput nature. This advantage is particularly useful for urgent diagnosis of or screening for infectious diseases that require immediate treatment and/or infectious control such as diseases caused by highly virulent and contagious agents and infection in critically ill patients or immunocompromised hosts. However, this technique should not replace but may serve as a complement conventional testing in clinical laboratories. The future application of microarrays in routine investigation of the infectious diseases depends on the development of more cost-effective protocols and equipments, more robust and simplified formats, easier or user friendly step of data analysis, and the adequate evaluation of their performance (efficacy) and convenience (efficiency) compared with other molecular methods. Hence, it is necessary to develop more simple assays that could be performed for all diagnostic laboratories.

In conclusion, microarrays will serve as a powerful tool that can be applied in several aspects of infectious diseases including diagnosis, identification of microbial and host factors related to prognosis and response to treatment, screening for antimicrobial resistance, epidemiological investigation, evolution study of microbes, identification of drug targets and development of new antimicrobials, and vaccine design. Thus, the microarray technology will revolutionize infectious disease practices in the future. However, clinicians must learn to use it effectively and appropriately and realize its limitation. The obtained accurate results, it needs to be interpreted in conjunction with other clinical and laboratory data.

## References

1. Schena M, Shalon D, Davis RW, Brown PO. Quantitative monitoring of gene expression patterns with a complementary DNA microarray. *Science* 1995 Oct 20; 270(5235): 467-70
2. Schena M, Heller RA, Theriault TP, Konrad K, Lachenmeier E, Davis RW. Microarrays: biotechnology's discovery platform for functional genomics. *Trends Biotechnol* 1998 Jul; 16(7): 301-6
3. Heller MJ. DNA microarray technology: devices, systems, and applications. *Annu Rev Biomed Eng* 2002; 4: 129-53
4. Dufva M. Fabrication of high quality microarrays. *Biomol Eng* 2005 Dec; 22(5-6): 173-84
5. Jaluria P, Konstantopoulos K, Betenbaugh M, Shiloach J. A perspective on microarrays: current applications, pitfalls, and potential uses. *Microb Cell Fact* 2007; 6: 4
6. Osin P, Shipley J, Lu YJ, Crook T, Gusterson BA. Experimental pathology and breast cancer genetics: new technologies. *Recent Results Cancer Res* 1998; 152: 35-48
7. Bloom G, Yang IV, Boulware D, Kwong KY, Coppola D, Eschrich S, Quackenbush J, Yeatman TJ. Multi-platform, multi-site, microarray-based human tumor classification. *Am J Pathol* 2004 Jan; 164(1): 9-16
8. Pollack JR. A perspective on DNA microarrays in pathology research and practice. *Am J Pathol* 2007 Aug; 171(2): 375-85
9. Bryant PA, Venter D, Robins-Browne R, Curtis N. Chips with everything: DNA microarrays in infectious diseases. *Lancet Infect Dis* 2004

- Feb; 4(2): 100 -11
10. Campbell CJ, Ghazal P. Molecular signatures for diagnosis of infection: application of microarray technology. *J Appl Microbiol* 2004; 96(1): 18-23
  11. Chen T. DNA microarrays—an armory for combating infectious diseases in the new century. *Infect Disord Drug Targets* 2006 Sep; 6(3): 263-79
  12. Cummings CA, Relman DA. Using DNA microarrays to study host-microbe interactions. *Emerg Infect Dis* 2000 Sep-Oct; 6(5): 513-25
  13. Kato-Maeda M, Gao Q, Small PM. Microarray analysis of pathogens and their interaction with hosts. *Cell Microbiol* 2001 Nov; 3(11): 713-9
  14. Hossain H, Tchatalbachev S, Chakraborty T. Host gene expression profiling in pathogen-host interactions. *Curr Opin Immunol* 2006 Aug; 18(4): 422-9
  15. Chizhikov V, Rasooly A, Chumakov K, Levy DD. Microarray analysis of microbial virulence factors. *Appl Environ Microbiol* 2001 Jul; 67(7): 3258-63
  16. Chemlal K, Portaels F. Molecular diagnosis of nontuberculous mycobacteria. *Curr Opin Infect Dis* 2003 Apr; 16(2): 77-83
  17. Seifarth W, Spiess B, Zeilfelder U, Speth C, Hehlmann R, Leib-Mosch C. Assessment of retroviral activity using a universal retrovirus chip. *J Virol Methods* 2003 Sep; 112(1-2): 79-91
  18. Sengupta S, Onodera K, Lai A, Melcher U. Molecular detection and identification of influenza viruses by oligonucleotide microarray hybridization. *J Clin Microbiol* 2003 Oct; 41(10): 4542-50
  19. Lin B, Vora GJ, Thach D, Walter E, Metzgar D, Tibbetts C, Stenger DA. Use of oligonucleotide microarrays for rapid detection and serotyping of acute respiratory disease-associated adenoviruses. *J Clin Microbiol* 2004 Jul; 42(7): 3232-9
  20. Vora GJ, Meador CE, Bird MM, Bopp CA, Andreadis JD, Stenger DA. Microarray-based detection of genetic heterogeneity, antimicrobial resistance, and the viable but nonculturable state in human pathogenic *Vibrio* spp. *Proc Natl Acad Sci USA* 2005 Dec 27; 102(52): 19109-14
  21. Korimbocus J, Scaramozzino N, Lacroix B, Crance JM, Garin D, Vernet G. DNA probe array for the simultaneous identification of herpesviruses, enteroviruses, and flaviviruses. *J Clin Microbiol* 2005 Aug; 43(8): 3779 - 87
  22. Loy A, Bodrossy L. Highly parallel microbial diagnostics using oligonucleotide microarrays. *Clin Chim Acta* 2006 Jan; 363(1-2): 106 -19
  23. Cleven BE, Palka-Santini M, Gielen J, Meembor S, Kronke M, Krut O. Identification and characterization of bacterial pathogens causing bloodstream infections by DNA microarray. *J Clin Microbiol* 2006 Jul; 44(7): 2389-97
  24. Wiesinger-Mayr H, Vierlinger K, Pichler R, Kriegner A, Hirschl AM, Presterl E, Bodrossy L, Noehammer C. Identification of human pathogens isolated from blood using microarray hybridisation and signal pattern recognition. *BMC Microbiol* 2007; 7: 78
  25. Li H, McCormac MA, Estes RW, Sefers SE, Dare RK, Chappell JD, Erdman DD, Wright PF, Tang YW. Simultaneous detection and high-

- throughput identification of a panel of RNA viruses causing respiratory tract infections. J Clin Microbiol 2007 Jul; 45(7): 2105-9
26. Spiess B, Seifarth W, Hummel M, Frank O, Fabarius A, Zheng C, Morz H, Hehlmann R, Buchheidt D. DNA microarray-based detection and identification of fungal pathogens in clinical samples from neutropenic patients. J Clin Microbiol 2007 Nov; 45(11): 3743-53
27. Palacios G, Quan PL, Jabado OJ, Conlan S, Hirschberg DL, Liu Y, Zhai J, Renwick N, Hui J, Hegyi H, et al. Panmicrobial oligonucleotide array for diagnosis of infectious diseases. Emerg Infect Dis 2007 Jan; 13(1): 73-81
28. Cho JC, Tiedje JM. Bacterial species determination from DNA-DNA hybridization by using genome fragments and DNA microarrays. Appl Environ Microbiol 2001 Aug; 67(8): 3677-82
29. Liu WT, Mirzabekov AD, Stahl DA. Optimization of an oligonucleotide microchip for microbial identification studies: a non-equilibrium dissociation approach. Environ Microbiol 2001 Oct; 3(10): 619-29
30. Malloff CA, Fernandez RC, Lam WL. Bacterial comparative genomic hybridization: a method for directly identifying lateral gene transfer. J Mol Biol 2001 Sep 7; 312(1): 1-5
31. Wang D, Urisman A, Liu YT, Springer M, Ksiazek TG, Erdman DD, Mardis ER, Hickenbotham M, Magrini V, Eldred J, et al. Viral discovery and sequence recovery using DNA microarrays. PLoS Biol 2003 Nov; 1(2): E2
32. Volokhov D, Chizhikov V, Chumakov K, Rasooly A. Microarray-based identification of thermophilic *Campylobacter jejuni*, *C. coli*, *C. lari*, and *C. upsaliensis*. J Clin Microbiol 2003 Sep; 41(9): 4071-80
33. Volokhov D, Pomerantsev A, Kivovich V, Rasooly A, Chizhikov V. Identification of *Bacillus anthracis* by multiprobe microarray hybridization. Diagn Microbiol Infect Dis 2004 Jul; 49(3): 163-71
34. Kessler N, Ferraris O, Palmer K, Marsh W, Steel A. Use of the DNA flow-thru chip, a three-dimensional biochip, for typing and subtyping of influenza viruses. J Clin Microbiol 2004 May; 42(5): 2173-85
35. Klaassen CH, Prinsen CF, de Valk HA, Horrevorts AM, Jeunink MA, Thunnissen FB. DNA microarray format for detection and subtyping of human papillomavirus. J Clin Microbiol 2004 May; 42(5): 2152-60
36. Kostrzynska M, Bachand A. Application of DNA microarray technology for detection, identification, and characterization of food-borne pathogens. Can J Microbiol 2006 Jan; 52(1): 1-8
37. Zhu LX, Wang D, Zhang GB, Jiang D, Zhang ZW, Zhang Q, Mitchelson K, Cheng J. Development of a base stacking hybridization-based microarray method for rapid identification of clinical isolates. Diagn Microbiol Infect Dis 2007 Oct; 59(2): 149-56
38. Zhu LX, Zhang ZW, Wang C, Yang HW, Jiang D, Zhang Q, Mitchelson K, Cheng J. Use of a DNA microarray for simultaneous detection of antibiotic resistance genes among staphylococcal clinical isolates. J Clin Microbiol 2007 Nov; 45(11): 3514-21
39. Wang XW, Zhang L, Jin LQ, Jin M, Shen ZQ, An S, Chao FH, Li JW. Development and

- application of an oligonucleotide microarray for the detection of food-borne bacterial pathogens. *Appl Microbiol Biotechnol* 2007 Aug; 76(1): 225-33
40. Tembe W, Zavaljevski N, Bode E, Chase C, Geyer J, Wasieloski L, Benson G, Reifman J. Oligonucleotide fingerprint identification for microarray-based pathogen diagnostic assays. *Bioinformatics* 2007 Jan 1; 23(1): 5-13
41. Schoolnik GK. Microarray analysis of bacterial pathogenicity. *Adv Microb Physiol* 2002; 46: 1-45
42. Jansen A, Yu J. Differential gene expression of pathogens inside infected hosts. *Curr Opin Microbiol* 2006 Apr; 9(2): 138-42
43. Feezor RJ, Oberholzer C, Baker HV, Novick D, Rubinstein M, Moldawer LL, Pribble J, Souza S, Dinarello CA, Ertel W, et al. Molecular characterization of the acute inflammatory response to infections with gram-negative versus gram-positive bacteria. *Infect Immun* 2003 Oct; 71(10): 5803-13
44. Bosinger SE, Hosiawa KA, Cameron MJ, Persad D, Ran L, Xu L, Boulassel MR, Parenteau M, Fournier J, Rud EW, et al. Gene expression profiling of host response in models of acute HIV infection. *J Immunol* 2004 Dec 1; 173(11): 6858-63
45. Kash JC, Basler CF, Garcia-Sastre A, Carter V, Billharz R, Swayne DE, Przygodzki RM, Taubenberger JK, Katze MG, Tumpey TM. Global host immune response: pathogenesis and transcriptional profiling of type A influenza viruses expressing the hemagglutinin and neuraminidase genes from the 1918 pandemic virus. *J Virol*. 2004 Sep; 78(17): 9499-511
46. Sexton AC, Good RT, Hansen DS, D'Ombra MC, Buckingham L, Simpson K, Schofield L. Transcriptional profiling reveals suppressed erythropoiesis, up-regulated glycolysis, and interferon-associated responses in murine malaria. *J Infect Dis* 2004 Apr 1; 189(7): 1245-56
47. Andes D, Lepak A, Pitula A, Marchillo K, Clark J. A simple approach for estimating gene expression in *Candida albicans* directly from a systemic infection site. *J Infect Dis* 2005 Sep 1; 192(5): 893-900
48. Kim HS, Choi EH, Khan J, Roilides E, Francesconi A, Kasai M, Sein T, Schaufele RL, Sakurai K, Son CG, et al. Expression of genes encoding innate host defense molecules in normal human monocytes in response to *Candida albicans*. *Infect Immun* 2005 Jun; 73(6): 3714-24
49. Matussek A, Strindhall J, Stark L, Rohde M, Geffers R, Buer J, Kihlstrom E, Lindgren PE, Lofgren S. Infection of human endothelial cells with *Staphylococcus aureus* induces transcription of genes encoding an innate immunity response. *Scand J Immunol* 2005 Jun; 61(6): 536-44
50. Reghunathan R, Jayapal M, Hsu LY, Chng HH, Tai D, Leung BP, Melendez AJ. Expression profile of immune response genes in patients with Severe Acute Respiratory Syndrome. *BMC Immunol* 2005; 6: 2
51. Stintzi A, Marlow D, Palyada K, Naikare H, Panciera R, Whitworth L, Clarke C. Use of genome-wide expression profiling and mutagenesis to study the intestinal lifestyle of *Campylobacter jejuni*. *Infect Immun* 2005

- Mar; 73(3): 1797-810
52. Cortez KJ, Lyman CA, Kottlilil S, Kim HS, Roilides E, Yang J, Fullmer B, Lempicki R, Walsh TJ. Functional genomics of innate host defense molecules in normal human monocytes in response to *Aspergillus fumigatus*. Infect Immun 2006 Apr; 74(4): 2353-65
53. Fornek JL, Korth MJ, Katze MG. Use of functional genomics to understand influenza-host interactions. Adv Virus Res 2007; 70: 81-100
54. Simitopoulou M, Roilides E, Likartsis C, Ioannidis J, Orfanou A, Paliogianni F, Walsh TJ. Expression of immunomodulatory genes in human monocytes induced by voriconazole in the presence of *Aspergillus fumigatus*. Antimicrob Agents Chemother 2007 Mar; 51(3): 1048-54
55. Waddell SJ, Butcher PD. Microarray analysis of whole genome expression of intracellular *Mycobacterium tuberculosis*. Curr Mol Med 2007 May; 7(3): 287-96
56. Ramilo O, Allman W, Chung W, Mejias A, Ardura M, Glaser C, Wittkowski KM, Piqueras B, Banchereau J, Palucka AK, et al. Gene expression patterns in blood leukocytes discriminate patients with acute infections. Blood 2007 Mar 1; 109(5): 2066-77
57. Motomura K, Toyoda N, Oishi K, Sato H, Nagai S, Hashimoto S, Tugume SB, Enzama R, Mugewa R, Mutuluza CK, et al. Identification of a host gene subset related to disease prognosis of HIV-1 infected individuals. Int Immunopharmacol 2004 Dec 20; 4(14): 1829-36
58. Lin MT, Albertson TE. Genomic polymorphisms in sepsis. Crit Care Med 2004 Feb; 32(2): 569-79
59. Clementi M, Di Gianantonio E. Genetic susceptibility to infectious diseases. Reprod Toxicol 2006 May; 21(4): 345-9
60. Dhiman N, Bonilla R, O'Kane DJ, Poland GA. Gene expression microarrays: a 21st century tool for directed vaccine design. Vaccine 2001 Oct 12; 20(1-2): 22-30
61. Loch C, Antoine R, Raze D, Mielcarek N, Hot D, Lemoine Y, Mascart F. *Bordetella pertussis* from functional genomics to intranasal vaccination. Int J Med Microbiol 2004 Apr; 293(7-8): 583-8
62. de la Fuente J, Ayoubi P, Blouin EF, Almazan C, Naranjo V, Kocan KM. Anaplasmosis: focusing on host-vector-pathogen interactions for vaccine development. Ann N Y Acad Sci 2006 Oct; 1078: 416-23
63. Lesko LJ. Personalized medicine: elusive dream or imminent reality? Clin Pharmacol Ther 2007 Jun; 81(6): 807-16
64. Murphy DJ, Brown JR. Identification of gene targets against dormant phase *Mycobacterium tuberculosis* infections. BMC Infect Dis 2007; 7: 84
65. Hervas F. Chip-mediated techniques: how close are we to generalised use in the infectious disease clinic? Clin Microbiol Infect 2004 Oct; 10(10): 865-7